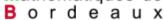


#### A plant pathogen multiscale problem dedicated to Chris Cosner

M. Langlais







SEGALEN











Courtesy from Pixels et Grains d'argent, and from INRA

#### Host-parasite system

- Vitis vinifera, vine stock & plot,
  - Spatial structures : leaves, plants, plots, no dispersal,
  - Seasonal variations : growing season, climate (temperature),
  - Highly anthropized system : wide diversity of cropping systems driving host dynamics (secondary shoots / leaves appearance),
  - No specific resistance of the host.

#### Host-parasite system

- Vitis vinifera, vine stock & plot,
  - **Spatial structures :** leaves, plant, plot scales, **no dispersal**,
  - Seasonal variations : growing season, climate (temperature),
  - Highly anthropized system : wide diversity of cropping systems driving host dynamics (secondary shoots / leaves appearance),
  - No specific resistance of the host.
- Erysiphe necator, powdery mildew,
  - Airborne dispersal of conidia (spores),
  - Sensitive to the climate : wind speed & direction (?),
  - Sensitive to the quality of its host : e.g., age of leaves, ...
  - Hard to detect and quantify at the vineyard level,
  - No reliable prevention tool : routine chemical sprays,
    - 30% world market in fungicides dedicated to vine, (F: 60 M€/year ),
  - Over-wintering in the bark of vine stocks & within dormant buds,

#### Outline : multi-scale problem

 (1) Architectural discrete 3D plant - pathogen model, discrete w.r.t. time & space, over a single growing season,

 $X(n+1) = F(X(n), n, \pi(n), \theta(n)), \quad n \ge 0.$ 

(2) Continuous aggregated plant - pathogen model, ODEs SEIRT-like, for a perennial plant,

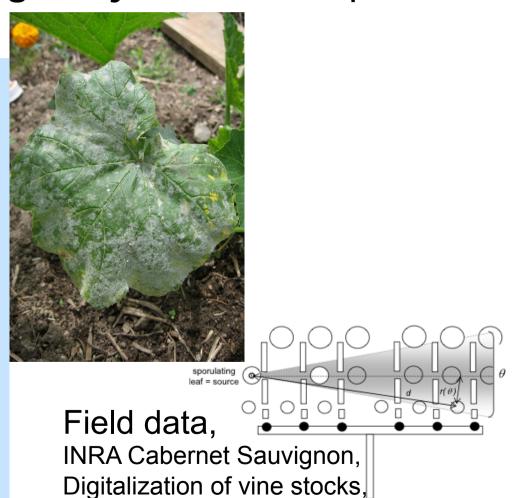
 $d_t X(t) = F^{year}(X(t), t, \pi(t)), \quad t \ge 0.$ 

(3) Spatially structured plot - pathogen model, RD - SEIR(T) like,

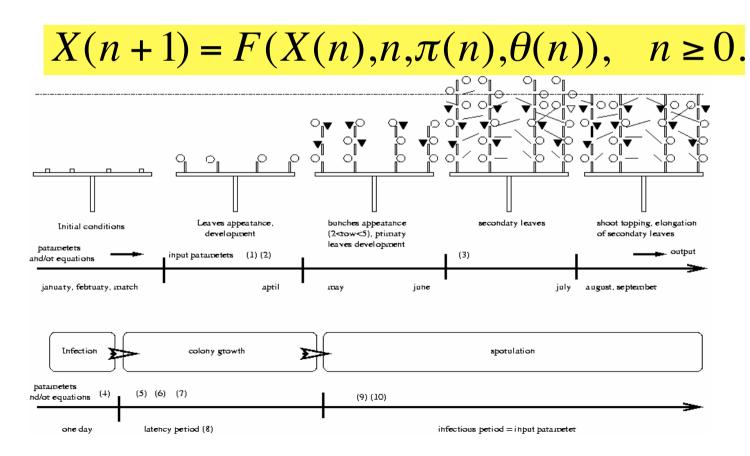
 $\partial_t U = \nabla \cdot (D(x,N)\nabla U) + V \cdot \nabla U + G^{year}(U,x,\pi(t)), \quad t \ge 0, x \in D.$ 

- Host dynamic: complex 3D architectural model,
- Appearance, growth, size, age, 3D localization and surface area of primary,
  - Leaves, internodes,
  - Shoots, grapes,
- Climatic scenario,
  - Daily basis,
- Secondary organs,
- Anthropization,
  - Shoot topping,

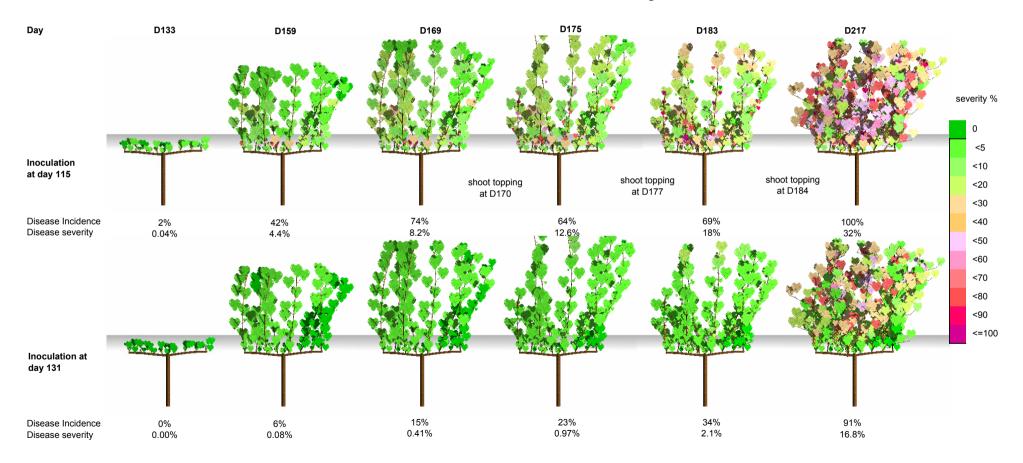
- Coupled host-pathogen dynamic : complex 3D architectural model,
- Primary infection :
  - Calendar date,
  - 3D localization on host,
- Lesion growth,
  - Latency period,
  - Spore production period,
  - Spore release,
- *Within* vine stock spore dispersal,
  - Speed of wind (daily),
- Secondary infections,



• Plant / pathogen dynamics for a growing season, complex 3D model, more than 30 parameters ...



• Scenario 1998, inoculation day : 115 vs 131,

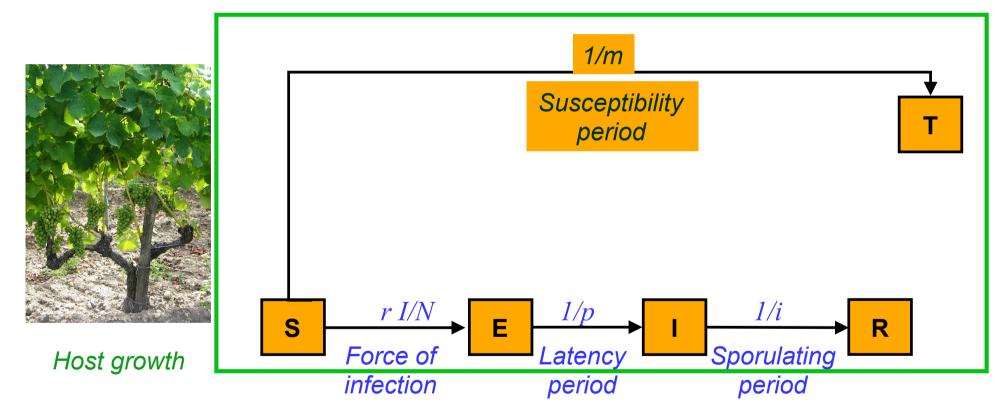


- PCA PLS analysis, Climatic scenarios and crop management do not impact the same variables,
  - Year (climate) is better correlated to early disease,
  - Crop management is more correlated to later disease,
- Sensitivity and elasticity analysis,
  - Age of leaves : susceptibility, ontogenic resistance,
  - Vigour of the host,
  - Sporulation,

IN?/

- Inoculation date,
- Temperature, dominant wind, ...
- Qualitative analysis ?

• SEIRT compartmental model w.r.t. leaf surface area,



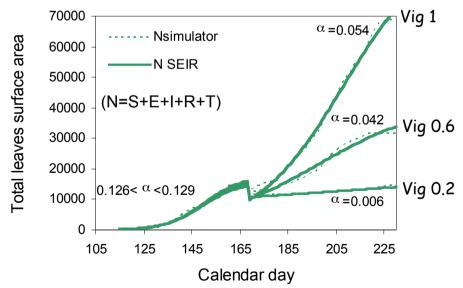
– incidence : r×S×I/N, Vanderplank, etc.

SEIRT model : Susceptible, Exposed,
 Infectious, Retired, T (on togenic resistance),

$$\begin{cases} d_t S = \Lambda - rI \frac{S}{N} - \frac{1}{m}S \\ d_t E = +rI \frac{S}{N} - \frac{1}{p}E \\ d_t I = + \frac{1}{p}E - \frac{1}{i}I \\ d_t R = + \frac{1}{i}I \\ d_t R = + \frac{1}{i}I \\ d_t T = + \frac{1}{m}S \end{cases} \Rightarrow d_t N = \Lambda = \alpha \left(1 - \frac{N}{K}\right)N.$$

– Find a set of parameters  $(r,\alpha,K)$  yielding consistent dynamics w.r.t. the discrete model ?

- « Aggregated » ODEs model fitted with discrete model,
  - Good fitting to aggregate time-dependent processes provided,
    ≠ sets of parameters (r,α,K) at shoot topping,
  - LSM : total leaf / diseased surface area,
  - Problem at shoot topping : status of remaining leaves ?

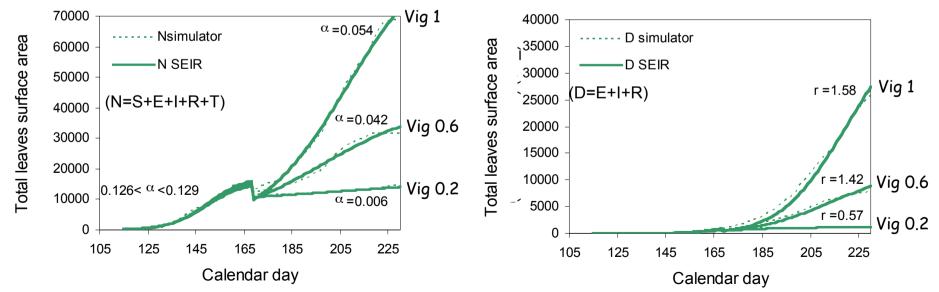


1998

- « Aggregated » ODEs model fitted with discrete model,
  - − Good fitting *to aggregate time-dependent processes* provided,  $\neq$  sets of parameters ( $r, \alpha, K$ ) at shoot topping,
  - LSM : total leaf / diseased surface area,

1998

– Problem at shoot topping : status of remaining leaves ?



- Qualitative analysis wrt standard epidemiology,
  - Computing the reproductive number  $R_0$ ,  $R_0 = r \times i$ ,
  - Computing the effective reproductive number  $R_{\text{effective}}$ ,
    - $\clubsuit \ R_{\text{effective}} = r \ i \ S(t) / N(t),$
    - ★  $t \rightarrow R_{\text{effective}}(t)$  « decreasing outside shoot-topping » ?

- Qualitative analysis wrt standard epidemiology,
  - Computing the reproductive number  $R_0$ ,  $R_0 = r^* i$ ,
  - Computing the effective reproductive number  $R_{\text{effective}}$ ,
    - $\clubsuit \ R_{\text{effective}} = r \ i \ S(t) / N(t),$
    - ★  $t \rightarrow R_{\text{effective}}(t)$  « decreasing outside shoot-topping » ?
  - Computing the asymptotic state in closed form (+ $\infty$  = winter)?
    - ♦ without ontogenic resistance (1/m = 0 or T = 0),

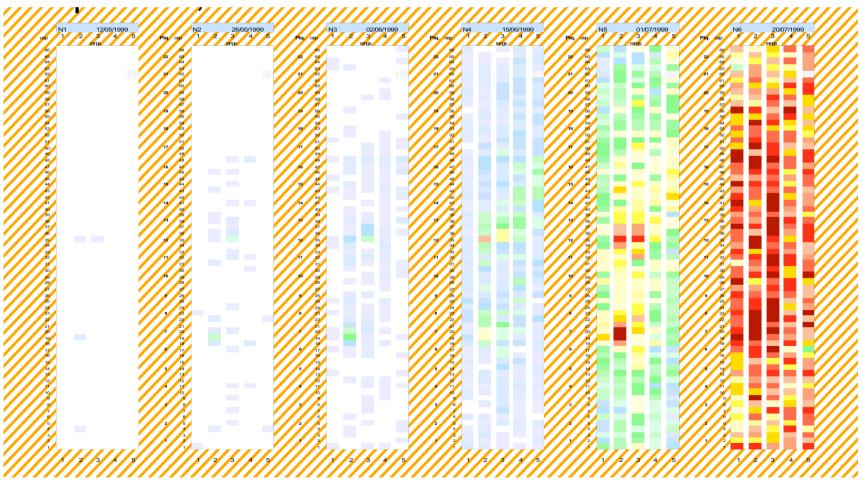
 $S = S^{infinity}, E = I = 0, R = R^{infinity}.$ 

✤ with ontogenic resistance,

$$S = 0$$
,  $E = I = 0$ ,  $R = R^{infinity}$ ,  $T = T^{infinity}$ .

#### Field data (1999), INRA Bordeaux

 Experimental plot with 5 rows & 66 vine stocks per row,



Pathogen dispersal : conceptual analysis.

Long and short distance dispersal type 1 : short alone, type 2 : both (coalescence), type 3 : (scaling effect) reduced short / large long.

#### Shigesada, N., Kawasaki. K.

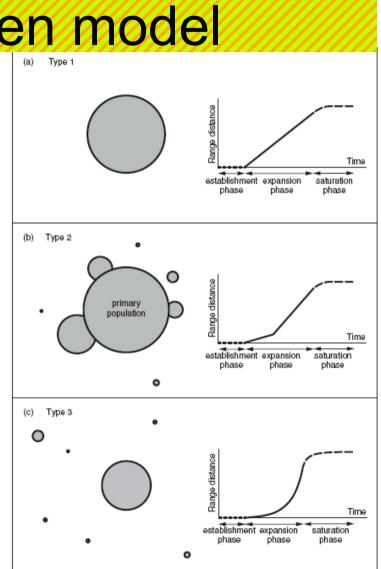
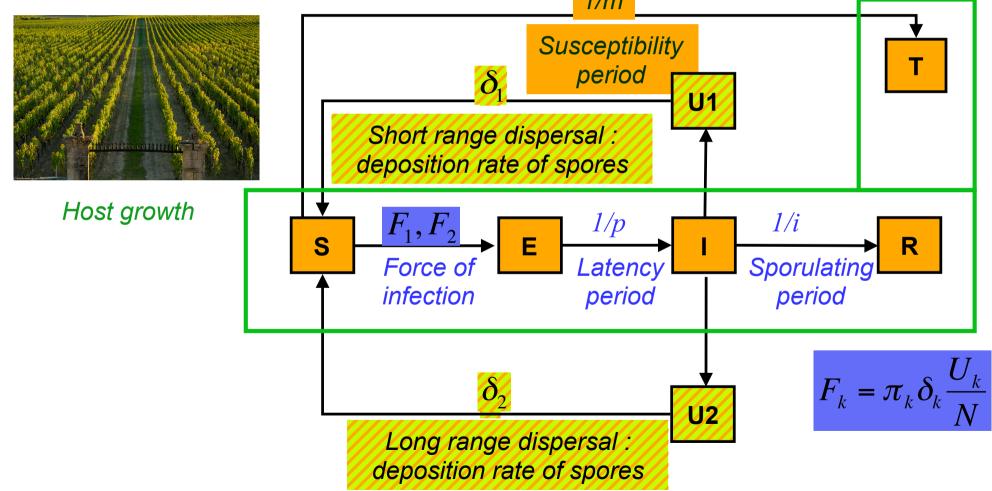


Figure 17.1 Three types of range expansions: (a) type 1, (b) type 2, and (c) type 3. For each type, the spatial pattern and range-versus-time curve are shown on the left and right, respectively. See text for detail. (Adapted from Shigesada & Kawasaki 1997, with permission of Oxford University Press.)

• SEIRT compartmental model over rows,



**R**eaction-**D**iffusion / SEIRT model :

(1) Reaction-Diffusion system for dispersal of spores,

 $\begin{cases} \partial_t U_1 = \nabla \cdot (d_1(x,N)\nabla U_1) - \delta_1 U_1 + \gamma f(N)I, \\ \\ \partial_t U_2 = \nabla \cdot (d_2(x,N)\nabla U_2) - V(x,t) \cdot \nabla U_2 - \delta_2 U_2 + \gamma (1 - f(N))I. \end{cases}$ 

 $\bullet$  U<sub>1</sub> & U<sub>2</sub>, short (plant scale) and long range dispersed spores,

- Fick's law diffusion,  $0 \le d_1(x,N) \le d_2(x,N)$ ,
- Convection term V(x,t), for average vs strong winds (?),
- ♦  $\delta_i$ , deposition rate of spores  $U_i$ , i=1,2,  $\delta_2 \leq \delta_1$ ,

 $\checkmark$   $\gamma$ , production rate of spores per infectious unit,

• f(N), % of spores dispersed at short range, Aylor (1999),

• Reaction-Diffusion / SEIRT model :

(2) SEIRT ODEs models over rows : production of spores by I and contamination of S by  $U_1$  and  $U_2$ 

 $\begin{cases} d_t S = \Lambda - (\pi_1 \delta_1 U_1 + \pi_2 \delta_2 U_2) \frac{S}{N} - \frac{1}{m} S \\ d_t E = + (\pi_1 \delta_1 U_1 + \pi_2 \delta_2 U_2) \frac{S}{N} - \frac{1}{p} E \end{cases} \qquad \begin{aligned} \pi_1 \gamma \approx r, \text{ODEs model}, \\ \pi_2 \leq \pi_1, \text{ inoculum eff.} \\ \pi_2 \leq \pi_1, \text{ inoculum eff.} \\ \end{cases} \\ \Rightarrow d_t N = \Lambda = \alpha \left(1 - \frac{N}{K}\right) N. \\ d_t R = + \frac{1}{i} I \\ d_t T = + \frac{1}{m} S \end{cases}$ 

- Basic model, R-D / SEIRT :
  - R-D system,

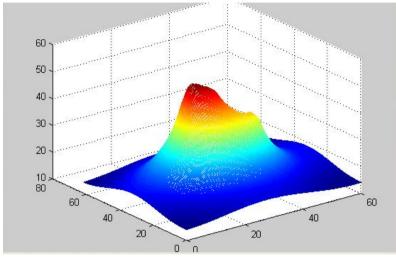
constant diffusivities, no convection term,

kinetic terms : homogeneous / heterogeneous,

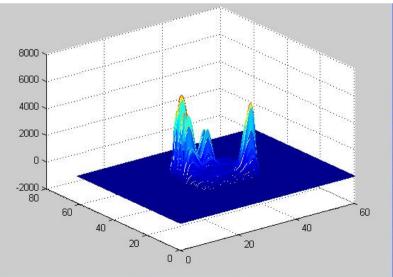
cf. Zawolek & Zadoks (1992), Vanderplank (1963),

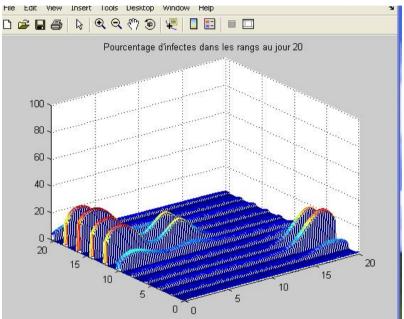
- Boundary conditions ?
  - *homogeneous Dirichlet conditions* set on a much larger domain than the spatial range of interest,
  - Mathematical analysis : IBVP.
  - Numerical experiments (Matlab interface).
    vs
- TW analysis.

# **Spatially structured** Spores : long range dispersal



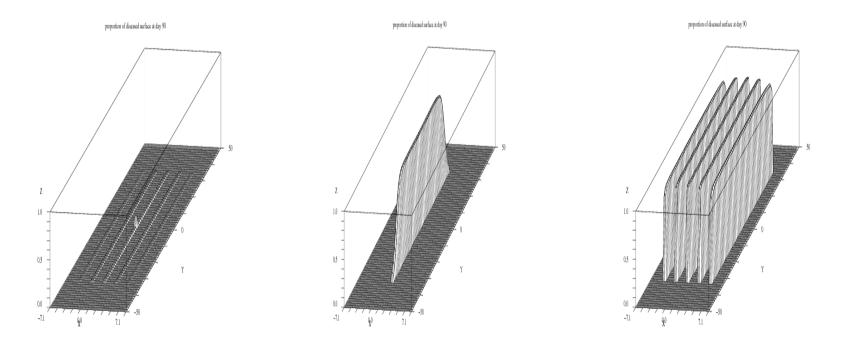
Spores : short range dispersal





#### Spatial spread of infection

- Proportion of disease lesion / dispersal, Day 90,
  - Long range dispersal only : weak uniform contamination,
  - Short range dispersal only : central row is contaminated,
  - Both : contamination spreads all over the five rows,



- Qualitative analysis :
  - Dimensionless model, R-D / SEIR (no T),

$$\frac{\partial U_1}{\partial t} = \Delta U_1 - \eta U_1 + \eta FI$$

$$\frac{\partial U_2}{\partial t} = d\Delta U_2 - \eta U_2 + \eta (1 - F)I$$

$$\frac{\partial S}{\partial t} = -R_0 (U_1 + U_2) S$$

$$\frac{\partial E}{\partial t} = +R_0 (U_1 + U_2)S - \frac{1}{\tau}E$$

$$\frac{\partial I}{\partial t} = \frac{1}{\tau} E - I$$

- Qualitative analysis (no T) :
  - Wave fronts, φ(x-c.t), 1D, pulses / infection, *linking*,
    - Pre-infection state,

♦  $U_1 = U_2 = 0$ , S = K, E = I = R = 0.

• Post-infection state,

♦  $U_1 = U_2 = 0$ ,  $S = K^{infinity}$ , E = I = 0,  $R = R^{infinity}$ .

- When  $R_0 > 1$ , minimal speed TW,  $c^*$ ,
  - implicit closed form (K<sup>infinity</sup>),
  - $K^{infinity}$  a root of,  $h = 1 e^{-R_0 h}$

• The value of c\* is given by solving (F<1) the characteristic equation:

Find 
$$(\lambda^*, c^*)$$
 such that  $0 < \lambda^* < \frac{c + \sqrt{c^2 + 4d\eta}}{2d}$  and  $\begin{cases} \psi(\lambda^*, c^*) = 1 \\ \frac{\partial \psi}{\partial \lambda}(\lambda^*, c^*) = 0 \end{cases}$ 

with

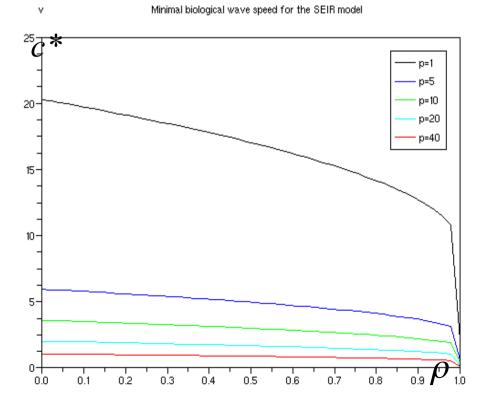
$$\psi_{SEIR}(\lambda,c) = \frac{R_0}{(1+c\lambda)(1+c\tau\lambda)} \left(\frac{\eta F}{\eta+c\lambda-\lambda^2} + \frac{\eta(1-F)}{\eta+c\lambda-d\lambda^2}\right)$$

- Dimensionless minimal speed, c\*,
  - Assuming :  $\delta_2 = \delta_1$  ,  $\pi_2 = \pi_1$  ,
  - $F(N)=\rho$ , % of short range dispersed spores,

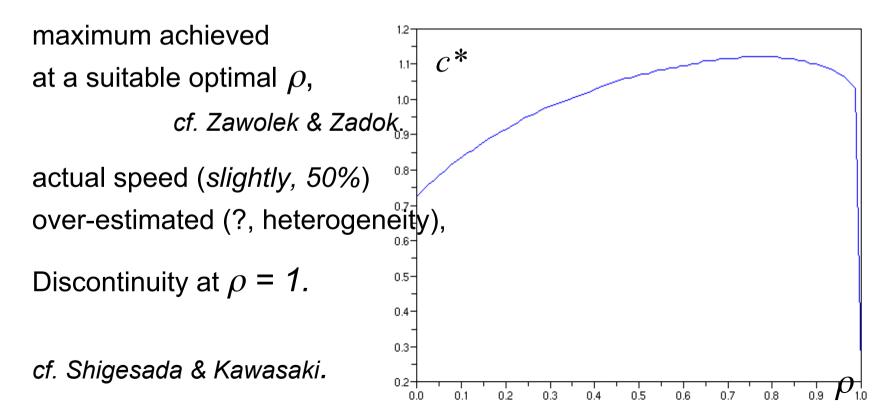
Decreasing w.r.t.  $\rho$ .

Discontinuity at  $\rho$  = 1.

cf. Shigesada & Kawasaki.



- Dimensionless minimal speed, c\*,
  - Assuming :  $\delta_2 < \delta_1$  ,  $\pi_2 < \pi_1$  ,
  - $F(N)=\rho$ , % of short range dispersed spores,



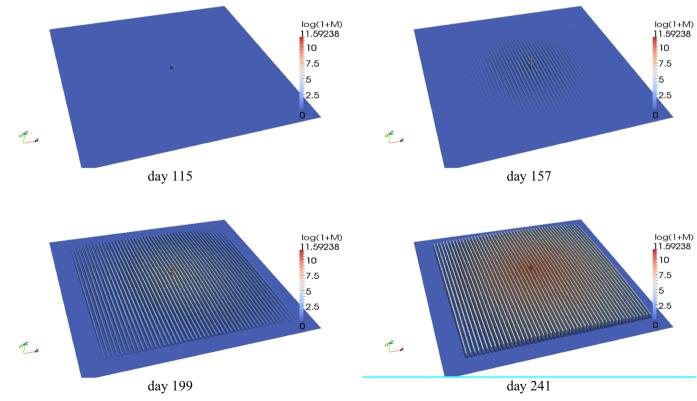
- 2D spatio-temporal periodical WF collaborative work A. Ducrot & H. Matano,
- *in which direction is the speed faster ?* numerics : along rows,

VS

data : across rows, but ...

 Numerical work in progress (turbulence, nonlinear diffusion, architecture & f(N), ...).

- 2D spatio-temporal dynamics,
- Nonlinear short range dispersal, data fitting ?



#### Further developments ...

- Collecting data and calibrating 2D and 3D models, including transport,
  - no known markers for *Erysiphe necator*, powdery mildew, compared to pollen dispersal,
  - Secondary infections from alien sources ...
- Control problems using fungicides,
  - when ?
  - coupling with shoot topping driving host dynamics ?
  - reducing or delaying the pathogen speed of propagation if coupled to ontogenic resistance ?

- Spatially periodic spatial plot,
  - reference cell, Y, "a single plant" (short range scale),
  - small parameter,  $\varepsilon > 0$ ,
  - spatial domain,  $\Omega$ ,

a set of cells of size  $\mathcal{E}^*Y$ ,



- available vegetal tissue density, function  $\chi$ , period *Y*,  $\chi(x,t,x/\varepsilon)$ .
- Simpler to handle as  $\varepsilon \rightarrow 0$  ?
  - spatially periodic: ill-conditioned



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